## Structure of the Human Reovirus Virion at 9.6Å Resolution

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Reovirus (family *Reoviridae*) is a large, icosahedral dsRNA virus with a diameter of ~850Å and a molecular mass of 129.5 MDa.[1] Reovirus virions are composed of eight proteins ( $\sigma$ 1,  $\sigma$ 2,  $\sigma$ 3,  $\mu$ 1,  $\mu$ 2,  $\lambda$ 1,  $\lambda$ 2,  $\lambda$ 3) and ten genome segments, which also code for three non-structural proteins ( $\mu$ NS,  $\sigma$ NS, and  $\sigma$ 1s). Six hundred copies each of  $\sigma$ 3 and  $\mu$ 1 are organized in the outer capsid in an incomplete, T=13 $\ell$  icosahedral lattice as 200  $\mu$ 1<sub>3</sub> $\sigma$ 3<sub>3</sub> heterohexamers. The  $\sigma$ 3 and  $\mu$ 1 proteins, which serve viral "protectin" and "penetrin" roles [2], are sequentially degraded by proteolysis inside endo/lysosomes. These events lead to release of the transcriptionally active reovirus core into the cytoplasm. The T=1 core, in addition to the genome, contains a shell (120  $\lambda$ 1, 150  $\sigma$ 2) and twelve pentameric turrets (60  $\lambda$ 2) and approximately 12-24 copies of the viral transcriptase ( $\mu$ 2,  $\lambda$ 3) which produce mRNA transcripts.[3] The  $\sigma$ 1 protein, which contains the receptor recognition function and confers tissue tropism, occurs as twelve trimers associated with the  $\lambda$ 2 turrets in virions.

Virions (serotype T3D) were embedded in vitreous ice and maintained at  $-176^{\circ}$ C as described.[4] Electron micrographs were recorded under low dose conditions (~24 electrons/Å<sup>2</sup>) in a Philips CM200 FEG microscope at a nominal magnification of 38,000×. Micrographs were digitized with a Zeiss PHODIS scanner with step size of 7 µm and bin-averaged to give 14 µm pixels (equivalent to 3.68Å at the specimen). Twenty-nine micrographs whose defocus ranged from 1.56 to 3.19 µm underfocus were selected for processing. Particle orientations and origins were determined using a model-based method.[5] The final three-dimensional reconstruction (FIG.1A,B), with corrections made to compensate for the effects of the microscope contrast transfer function, was computed from 3652 particles.[4] The distribution of particle orientations were <0.01) to allow computation of the reconstruction to the 9.6Å resolution limit of the data.[4] X-ray crystallographic structures of the core [6] and the µ1<sub>3</sub>\sigma3<sub>3</sub> heterohexamer [7] exhibited excellent agreement with the reconstructed density map. The program EMFIT [8] was used to accurately dock various components such as the µ1\sigma3 heterohexamer into the virion map.

Inspection of the density map revealed numerous rod-like features most of which could be ascribed to  $\alpha$ -helical secondary structural elements present in the X-ray structures of all five major structural proteins (FIG.1C-E). In addition, novel features present in the reconstructed density but not in the crystal structures were observed. For example, spokes of density emanate from and appear to interconnect the  $\mu$ 1 trimers at sites of local sixfold symmetry in the T=13 lattice (FIG.1F). One spoke projects away from each  $\mu$ 1 molecule and merges into an annular ring at the local sixfold axis. Recent evidence for the presence of stabilizing disulfide bonds in the outer capsid of orthoreovirus virions [2,7] is consistent with the observed hub-like structure. [9]

References:

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FIG.1. A. Surface representation of the reovirus T3D virion viewed along a 5-fold symmetry axis. One  $\lambda 2$  pentamer (black square) is shown at higher magnification in (B). C. Density projection of the T3D map at a radius of 338Å and viewed along a 3-fold axis (high density appears black). The square demarks a region that includes a small portion of three  $\mu_1$  subunits (enlarged in D). The circle identifies a portion of the P3 channel [1] and six surrounding  $\mu_1$  subunits (also shown in F). D. Fit of the reovirus T1L  $\mu_1$  X-ray crystal structure [7] into the T3D density map. The portion of the map shown, a planar section near the region depicted in (C; square box), reveals that several long stretches of  $\alpha$ -helices in the  $\mu_1$  trimer fit nicely into rod-like densities in the reconstructed map. Only the C<sub> $\alpha$ </sub> backbone of the  $\mu_1$  X-ray structure is depicted. E. Same as (D) but only showing the X-ray structure. F. Magnified view (shaded surface representation) of a planar section centered about the P3 channel (see encircled region in C). A spoke structure, suspended inside the channel at a particle radius of ~334Å, appears to arise from the association of six density features that project from each of the six  $\mu_1$  subunits that form the channel. Scale bars = 200Å (A,C); 50Å (B,F); and 20Å (D,E).