CHM 165,265 / BIMM 162 / BGGN 262 Spring 2013

Lecture Slides

Jan 24, 2013

CHM 165,265 / BIMM 162 / BGGN 262 Spring 2013 Announcements for Jan 24, 2013

Reading assignment for Tuesday: Lecture notes pp.124-195

Note: much of this material will NOT be covered in lecture

'Virtual' homework: Answers to first 4 sets posted outside NSB 4-105

Recitation session: Tomorrow 5:00-6:00 pm in York 4080A

TEM facility tour: NEXT WEEK Jan 28,29 (check web site)

CHM 165,265 / BIMM 162 / BGGN 262 Winter 2013 3D Electron Microscopy of Macromolecules

TEM Facility Tour

Where: 1510 Bonner Hall basement

When: Mon Jan 28 and Tue Jan 29th

Check class web site for details on dates, times, and directions to facility

Attendance is optional but 5 pts extra credit towards final grade will be awarded



FEI Technai Sphera (200keV; LaB₆; LN₂)

To reserve and guarantee a time slot, email **nholson@ucsd.edu First come, first served.**

I.C CONTRAST AND IMAGE FORMATION KEY CONCEPTS FROM LECTURE #5

- TEM images do **NOT** give a **completely faithful** rendering of the density distribution (*i.e.* structure) of specimens
- Relationship between image and specimen is described by the contrast transfer function (CTF), which is characteristic of or influenced by:

Specific TEM used (Obj lens C_s);
 Conditions of imaging (defocus used);
 Specimen

 <u>Microscope</u> CTF arises from the objective lens focal setting <u>AND</u> from the spherical aberration in the objective lens

$$CTF(v) = -\left\{ \left(1 - F_{amp}^{2}\right)^{\frac{1}{2}} \cdot \sin(\chi(v)) + F_{amp} \cdot \cos(\chi(v)) \right\}$$

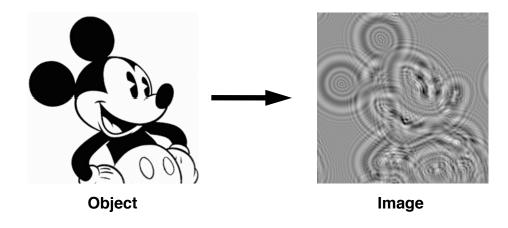
where $\chi(v) = \pi \lambda v^{2} \left(\Delta f - 0.5C_{s} \lambda^{2} v^{2}\right)$

I.C CONTRAST AND IMAGE FORMATION KEY CONCEPTS FROM LECTURE #5

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I.D ALIGNMENT/ADJUSTMENT OF THE TEM MORE CONCEPTS FROM LECTURE #5

- TEM alignment affects resolving power and convenience of operation
- Goal of alignment: make optical elements of TEM coaxial
- Principle for aligning any electromagnetic lens based on image rotations caused by fluctuating the current (or voltage) in lenses
- Small changes in objective lens current used to focus electron images

I.D ALIGNMENT/ADJUSTMENT OF THE TEM MORE CONCEPTS FROM LECTURE #5

Top Five Disturbances to Microscope Performance

- **Contamination** leads to astigmatism, drift, and decreased contrast (Use anticontaminator)
- Image drift and mechanical instabilities caused by instabilities in specimen holder, stage assembly, and specimen. (Measure drift rate)
- Electrical and magnetic instabilities (Use high voltage to distinguish)
- Image astigmatism ("Experts" able to correct this)
- Focal drift (Micro-discharges in gun?)



I.E OPERATION OF THE TEM MORE CONCEPTS FROM LECTURE #5



What happens as V is increased?

Specimen penetration increases Amplitude contrast **de**creases Resolution limit (diffraction) improves 0 0 Inelastic scattering decreases Chromatic aberration **de**creases (Radiation damage **de**creases • • Photographic emulsion efficiency decreases ••• CCD image quality **de**creases Electron gun **more sensitive** to vacuum quality ••• Electron gun brightness increases Screen phosphor efficiency increases

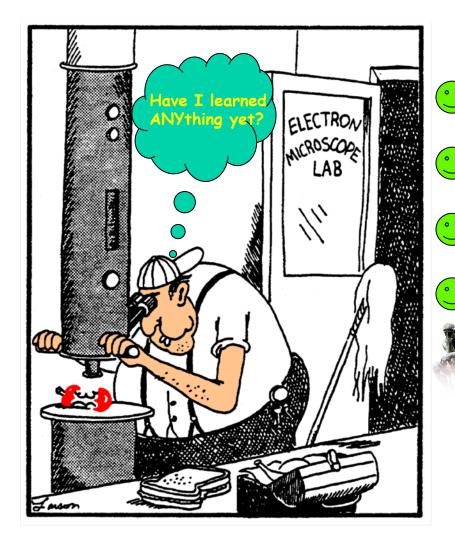


I.E OPERATION OF THE TEM MORE CONCEPTS FROM LECTURE #5

What happens as V is decreased?

Specimen penetration **de**creases Amplitude contrast increases Resolution limit (diffraction) gets worse 0 0 Inelastic scattering increases 0 0 Chromatic aberration increases • • Radiation damage increases Photographic emulsion efficiency increases CCD image quality increases Electron gun less sensitive to vacuum quality ••• Electron gun brightness **de**creases Screen phosphor efficiency **de**creases





TOPICS

- Principles of TEM Electrons, lenses and optics
 Design of TEM Components top to bottom
 Contrast and image formation Electron scattering from object
 Optimizing TEM performance Alignment assures 'best' images
 Operation of TEM "What do all these buttons do?"
 Other modes of TEM Many ways to 'observe' specimens
 - Specimen preparation for TEM Getting specimen ready
 - Radiation damage Less is better
 - 3D reconstruction

Specimen 3D structure from 2D images



Choice of accelerating voltage \star Choice of apertures Specimen stage/holder Choice of magnification Focusing Magnification calibration **Resolution tests** Image intensifiers/TV displays Microscope maintenance Photography (analog and digital)

p-Flasher Question

The contrast transfer function of the TEM is affected by which of the following?

- A. The defocus setting of the objective lens
- B. Spherical aberration in the objective lens
- C. The voltage and therefore wavelength of the imaging electrons
- D. The coherence of the electron beam
- E. All of the above

§ I: The Microscope

I.E Operation of the TEM

I.E.2 Choice of Apertures

- Condenser
- Objective

I.E.2 Choice of Apertures I.E.2.a <u>Condenser</u> Aperture

Bottom Line: Smaller is better.....up to a point.

Small apertures are best for high resolution imaging Produces a more coherent beam and better phase contrast

BUT: As condenser aperture size is reduced, fewer electrons are available to illuminate the specimen Harder to focus and correct astigmatism accurately Need for longer exposure times

I.E.2 Choice of Apertures

I.E.2.b <u>Objective</u> Aperture

Bottom Line: Smaller is also better.....up to a point.

Small apertures improve scattering/amplitude contrast The smaller the better, right?

Small apertures reduce spherical and chromatic aberrations The smaller the better, right?

BUT: As objective aperture size is reduced,

Diffraction limited resolution gets worse (Airy disk gets larger) Small apertures are harder to align and keep aligned, and are more sensitive to effects of contamination



Choice of accelerating voltage
 Choice of apertures

Bottom Line: Smaller is better.....up to a point.

Condenser: small apertures best for high resolution

Objective: small apertures improve scattering contrast and reduce spherical and chromatic aberrations

However:

Condenser: small apertures reduce illumination and hence make it harder to focus and correct astigmatism

Objective: small apertures lead to poorer diffraction limited resolution and are harder to align and keep aligned, and are more sensitive to the effects of contamination



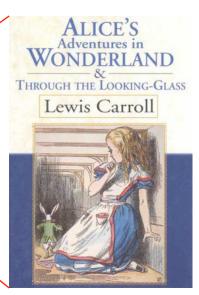
- \star Choice of accelerating voltage
- ★ Choice of apertures
- Specimen stage/holder
 Choice of magnification
 Focusing

Hands off the merchandise!!! Specimen grid: flat and secure (pp.91-92, lecture notes)

- Magnification calibration
- **Resolution tests**
- Image intensifiers/TV displays
- Microscope maintenance
- Photography (analog and digital)



- ★ Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
- Choice of magnification
 Focusing
 Magnification calibration
 Resolution tests
 Image intensifiers/TV displays
 Microscope maintenance
 - Photography (analog and digital)



- Is it appropriate?
- Field of view
- Radiation damage
- High resolution

I.E.4 Choice of Magnification

"Should I go higher or lower?"

Choice depends on the nature of the experiment

Many criteria for choosing a suitable magnification

I.E.4 Choice of Magnification

"Should I go higher or lower?"

Choose "low" (< 10,000X) to maximize field of view (*i.e.* get the BIG picture)

Choose "high" (> 30,000X) to maximize resolution captured in the image (*i.e.* to see fine details)

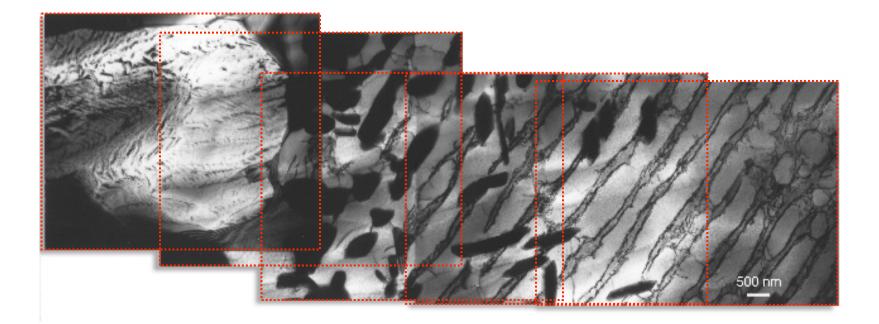
I.E.4 Choice of Magnification I.E.4.a Low Magnification (<10,000X)

- Necessary to capture a large field of view from a specimen in a single micrograph
- Montage: splicing together prints from several low magnification micrographs
- If very low magnifications are used, pincushion and barrel distortions could be significant and the prints won't match correctly

I.E.4.a Low Magnification (<10,000X)

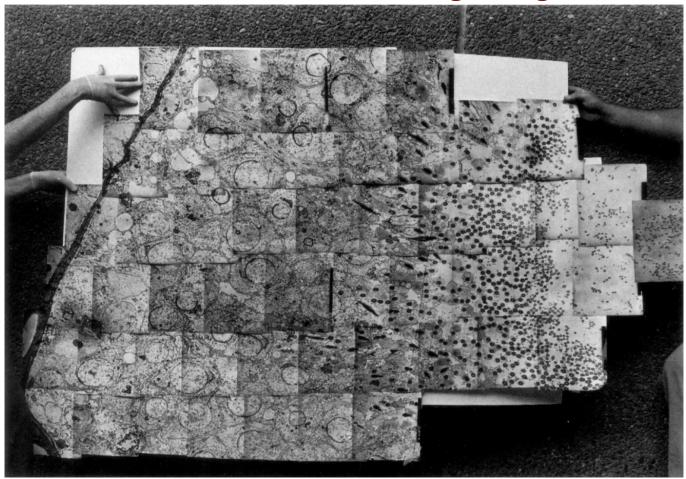
Use when need to capture a large field of view from a specimen in a single micrograph

Use if need to prepare a **montage**, where prints from several low magnification micrographs are spliced together



I.E.4.a Low Magnification (<10,000X)

Montage of several overlapping micrographs to show extended view of tissue at high magnification

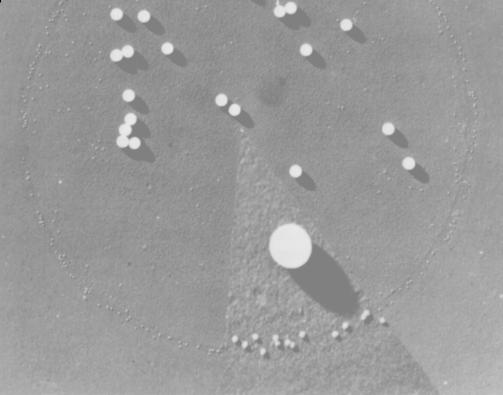


Slide not shown in class lecture

From Bozzola, 1st Ed., Fig. 8-14, p.233

I.E.4.b Statistical Studies

For statistical studies (particle counting) use the **lowest** magnification at which the particles can be correctly identified



Slide not shown in class lecture

From Hall, Fig. 10.65, p.359

I.E.4.c Radiation Damage

Use **lowest** possible magnification <u>and</u> illumination level when <u>searching the grid</u> for a suitable region of a radiation sensitive specimen

Use **lowest** possible magnification <u>and</u> illumination level when <u>recording an image</u> of the desired area **but high** enough to capture desired level of detail



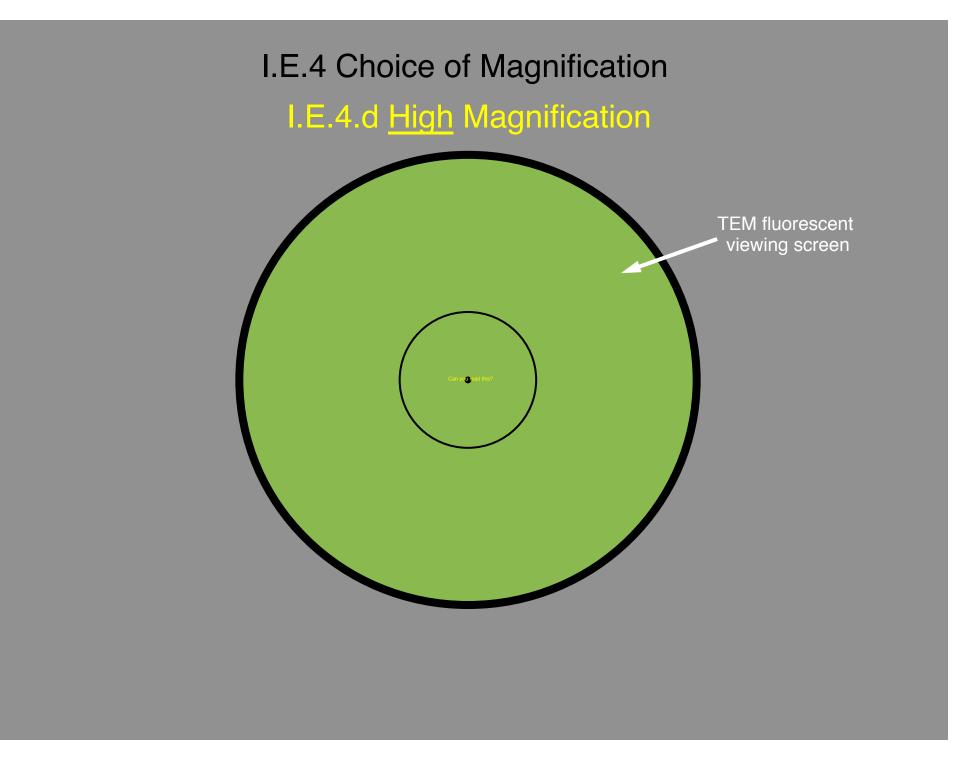
I.E.4 Choice of Magnification I.E.4.d <u>High</u> Magnification

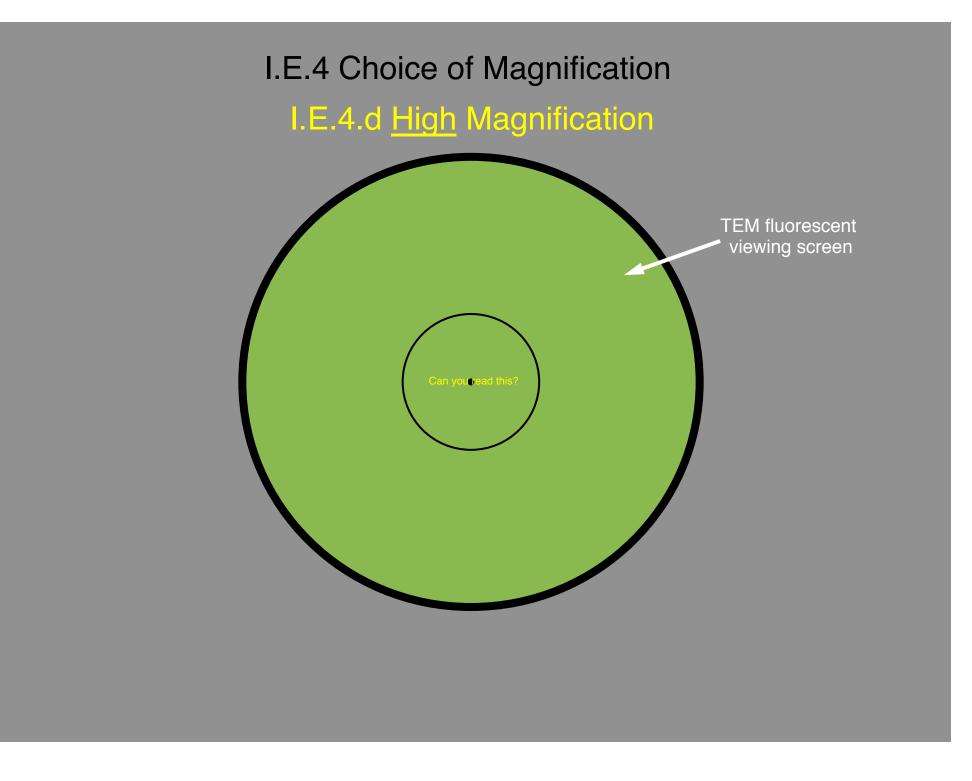


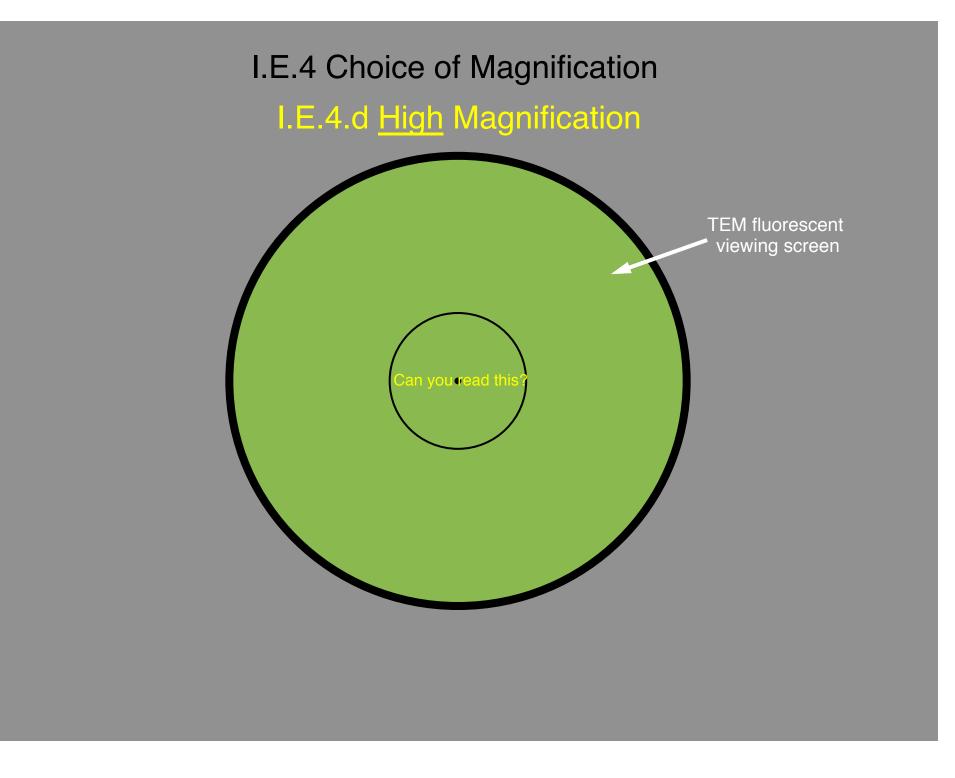
"High" magnification required to achieve highest resolution images (*i.e.* capture the finest specimen details)

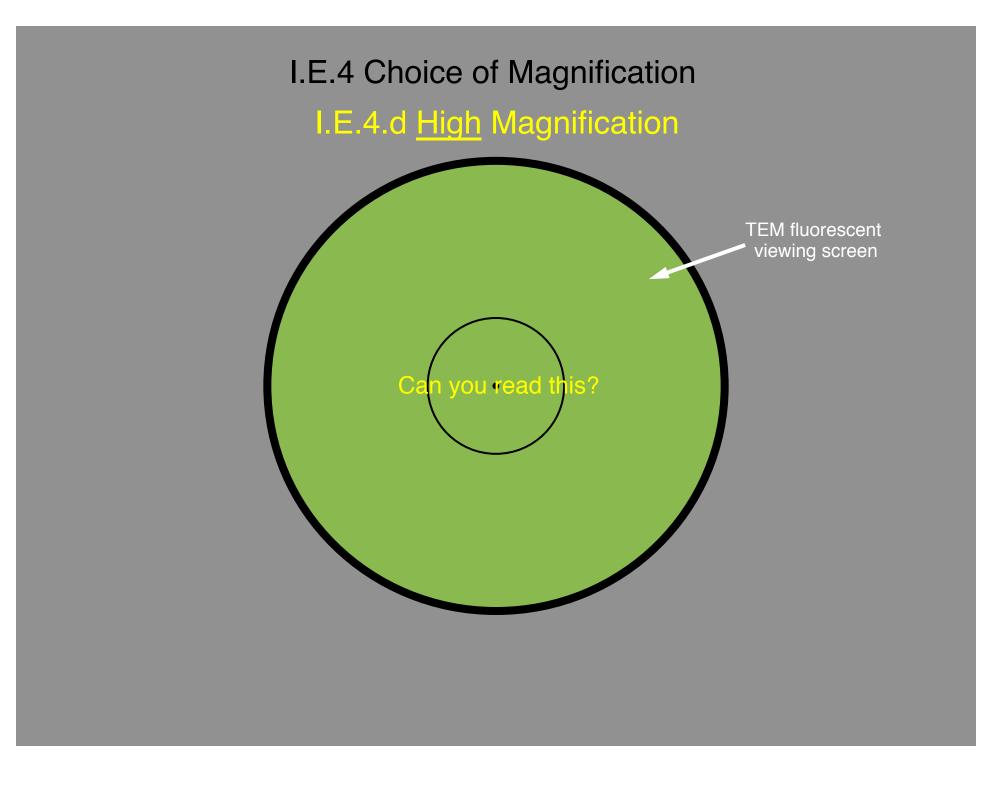
Excessive ("empty") magnification:

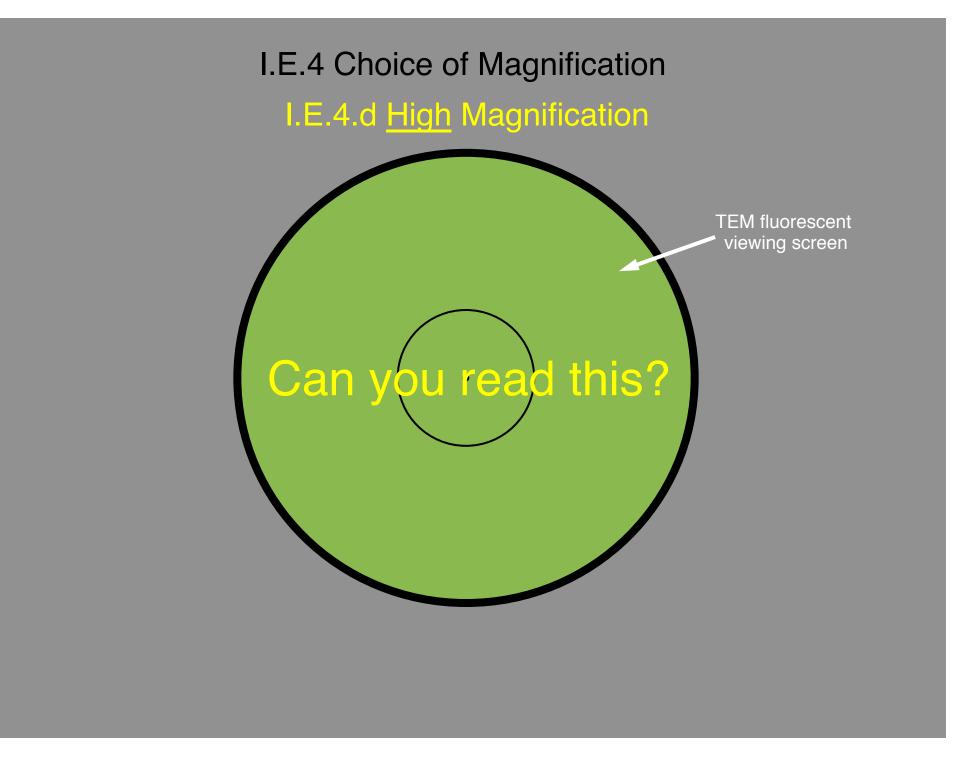
Leads to unnecessary **radiation damage** because higher doses are needed to expose a photographic emulsion or CCD/DDD camera

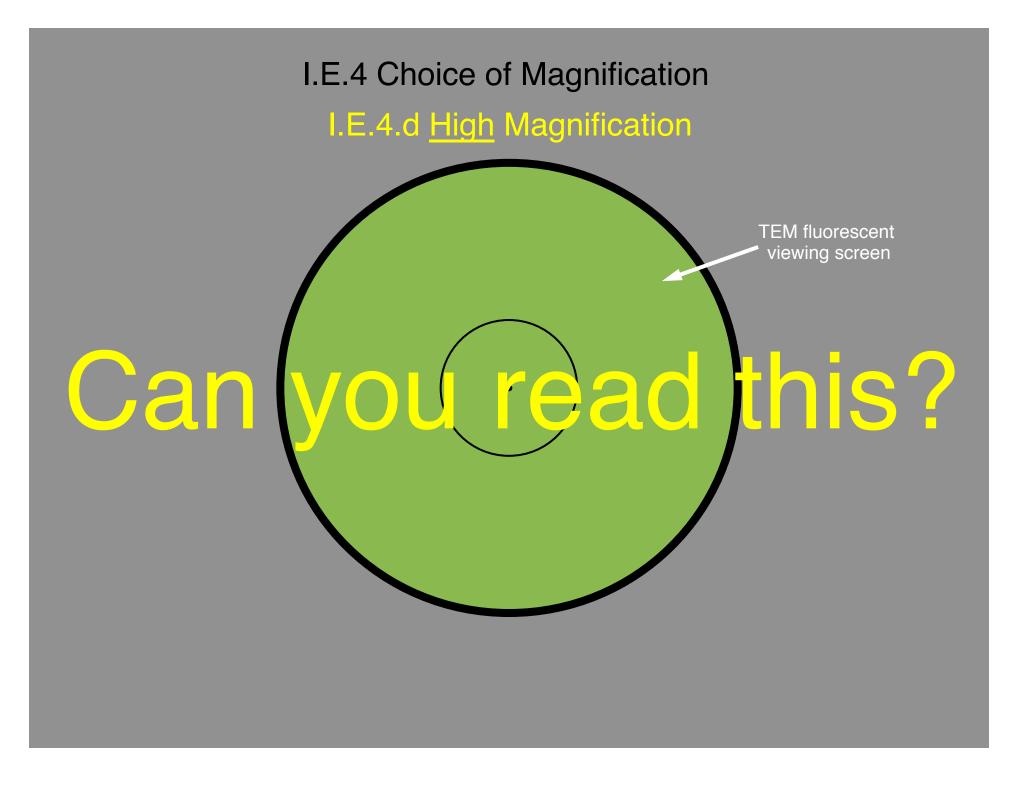


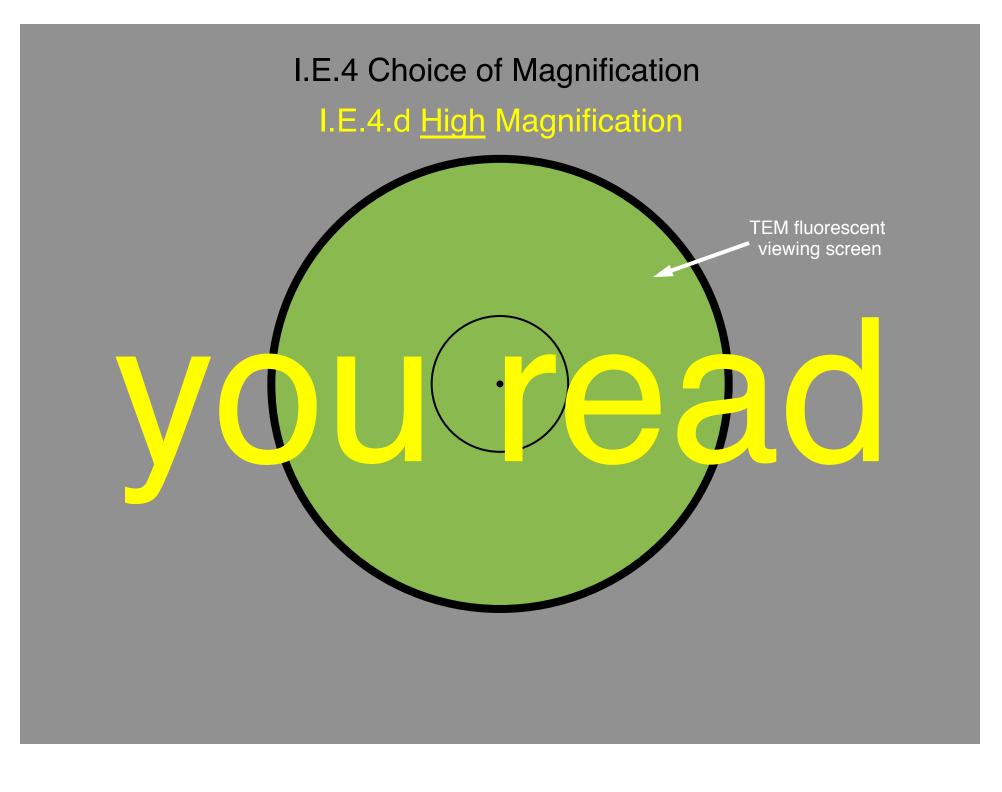


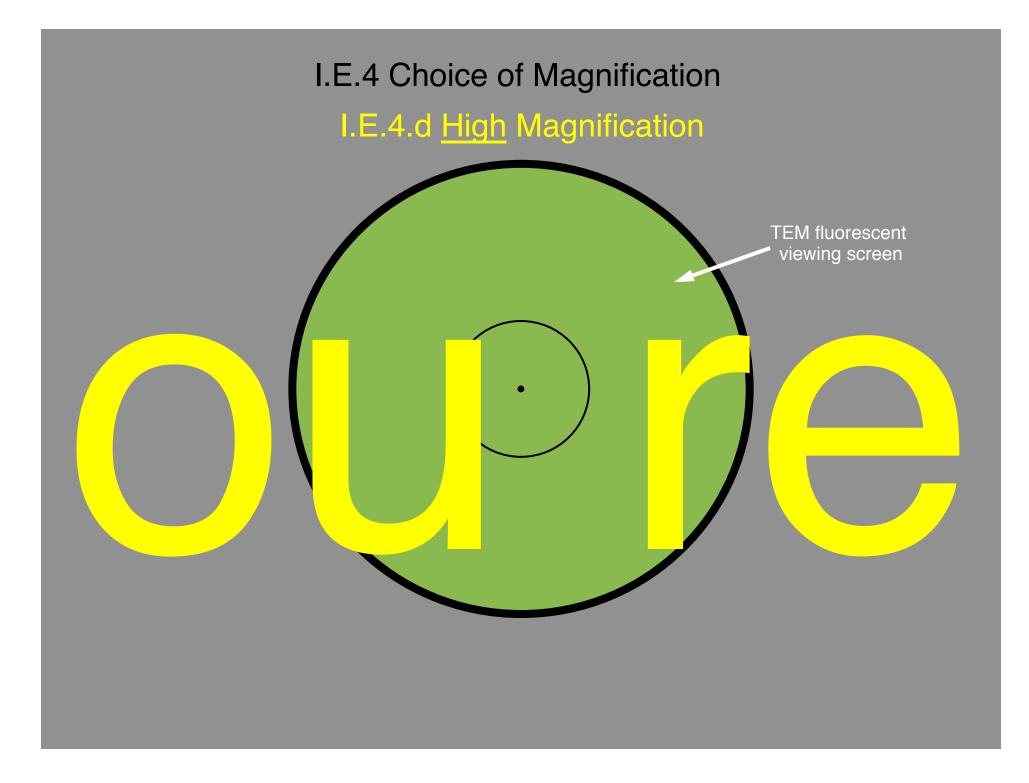




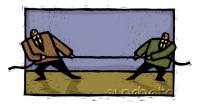








I.E.4.d High Magnification



Resolution of photographic medium (or pixel size of CCD or DDD camera) ultimately determines the **maximum** magnification needed to record details at a *predetermined or estimated* resolution

Photographic emulsions can resolve image details <u>AT LEAST</u> as small as 20 μ m

Theoretical (*i.e.* potential) resolution of object details captured in a photographic image depends on image magnification according to:

Potential resolution = $\frac{20 \ \mu m (`pixel' size)}{magnification}$

NOTE: This is a <u>conservative</u> estimate since good photographic films can resolve details as small as 5–10 µm.

I.E.4.d High Magnification



<u>Potential</u> resolution of object detail on film = 20 μ m/magnification

Magnification	Potential Resolution at Object
2,000	10.0 nm
20,000	1.0 nm
50,000	0.4 nm
100,000	0.2 nm

Take home message:

Given the limited resolution that can be obtained in images of biological specimens, most "typical" microscopy is performed at magnifications **much higher than necessary** (and restricts specimen field of view)

I.E.4 Choice of Magnification I.E.4.d <u>High Magnification</u>



Rule of Thumb:

Select **lowest** magnification consistent with required resolution and the recording medium used

NOTE: Each time magnification is doubled, a 4-fold increase in beam intensity is required at the <u>specimen</u> to maintain the same intensity level in the image



- ★ Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
- Choice of magnification
 Focusing
 Magnification calibration
 Resolution tests
 Image intensifiers/TV displays
 - Microscope maintenance
 - Photography (analog and digital)



- ★ Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
- \star Choice of magnification
- \Rightarrow Focusing
 - Magnification calibration
 - Resolution tests



- Image intensifiers/TV displays Microscope maintenance
- Photography (analog and digital)

p-Flasher Question

At which one of the following objective lens focal settings does interference contrast complement aperture contrast?

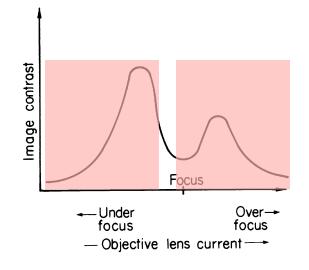
A. Slight under focus

- B. Slight over focus
- C. Near focus
- D. True focus
- E. Exact focus



I.E.5 Focusing

- Desired focus setting is set by making small changes in objective lens current
- True / near / exact / dead focus: condition where, ideally, no Fresnel fringes will form at an image point
- Slight degree of underfocusing gives optimum results

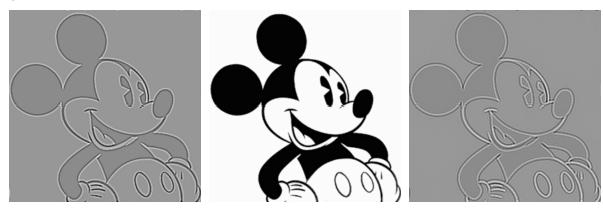


From Meek, 2nd ed., Fig. 5.3, p.100



I.E.5 Focusing

- Desired focus setting is set by making small changes in objective lens current
- True / near / exact / dead focus: condition where, ideally, no Fresnel fringes will form at an image point
- Slight degree of underfocusing gives optimum results
 - Here, aperture contrast will be **enhanced** with interference contrast (*i.e.* they work TOGETHER)

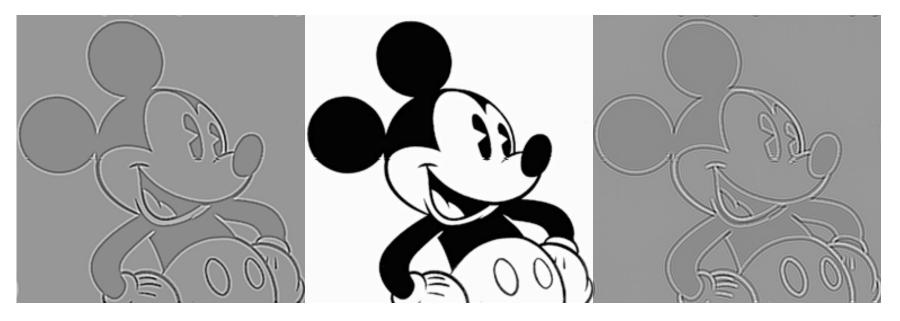


I.E.5 Focusing



Slight degree of underfocusing gives optimum results

Here, aperture contrast will be **enhanced** with interference contrast (*i.e.* they work TOGETHER)



Slight underfocus (5% Amplitude, 95% Phase)

In focus (100% Amplitude)

Slight overfocus (5% Amplitude, 95% Phase)

I.E.5 Focusing I.E.5.a Focusing at Low Magnification (< 15,000X)



Primary Methods

1. Wobbler

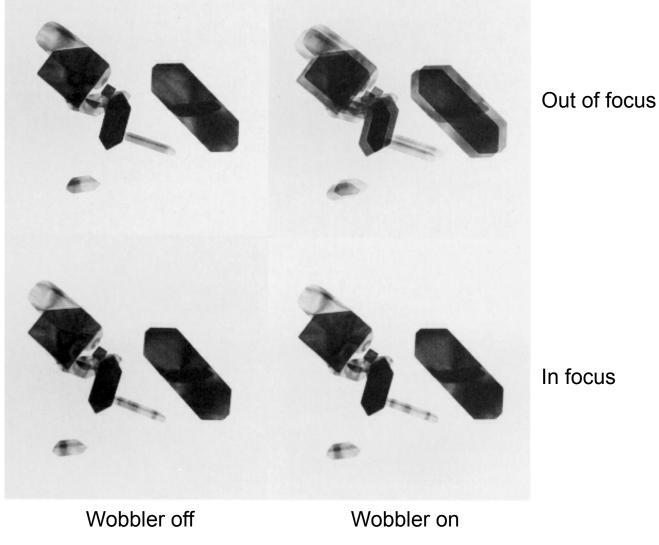
2. Minimum Contrast

(see p.93 of lecture notes)

I.E.5 Focusing I.E.5.a Focusing at Low Magnification (< 15,000X)



Focusing with a Wobbler Aid



From Watt, Fig. 2.11b, p. 30

I.E.5 Focusing



I.E.5.a Focusing at Low Magnification (< 15,000X)

2. Minimum Contrast Method

(see p.93 of lecture notes)

- A. Withdraw objective aperture
- B. Focus objective lens to minimize image phase contrast (i.e. try to make image "disappear")
- C. Reinsert objective aperture and record image

Not the most effective or recommended way to focus!

1) Objective aperture may become misaligned

2) Too crude for careful imaging esp. at high resolution

I.E.5 Focusing



I.E.5.a Focusing at Low Magnification (< 15,000X)

Primary Methods

1. Wobbler

2. Minimum Contrast

I.E.5.b <u>High</u> Magnification Focusing (usually > 30,000X)

Primary Method

Forget the specimen; use the support film!

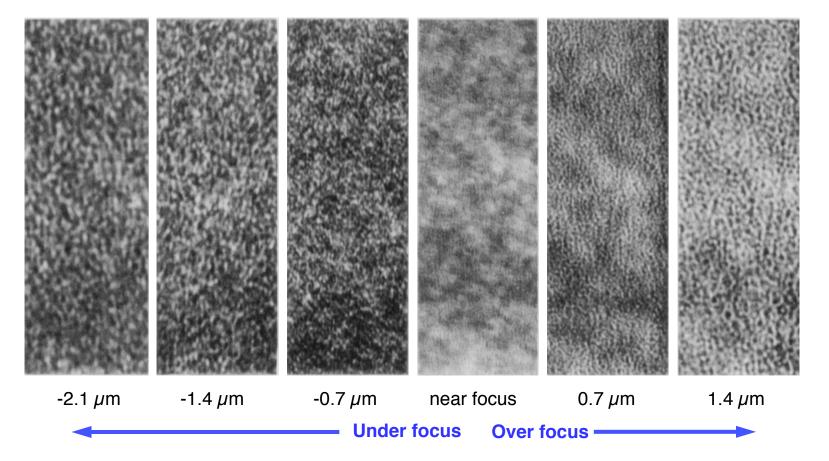


Forget the specimen; use the support film!

- Focus image based on the appearance of the support film ("sub-structure" seen is mainly just phase contrast Fresnel fringes)
- Best NOT to judge proper focus based merely on the appearance of a specimen, especially a new or unfamiliar one

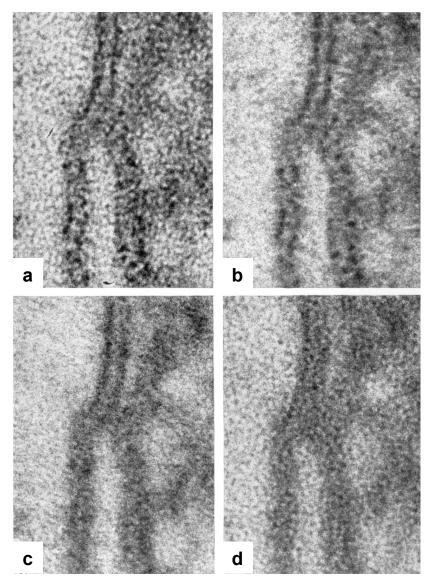
Support film is a predictable (well-defined, well-behaved) "specimen" with a characteristic appearance at near focus settings

Through Focus Series: Thin Carbon Film Lightly Shadowed with Platinum

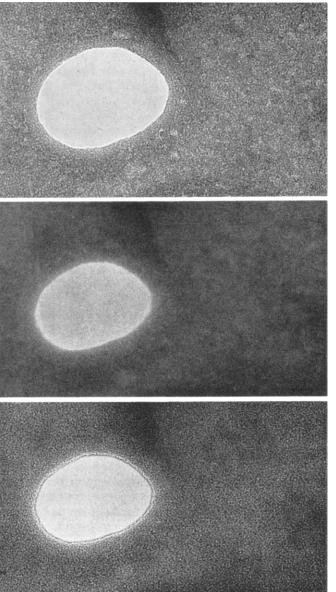


Phase contrast effects on image of stained mitochondrial outer membrane in thin section

- a) Under focus
- b) Close to focus
- c) Slightly over focused
- d) Over focused



From Sjostrand, Fig. IV.13, p. 109



Hole in a thin carbon film

Under focus

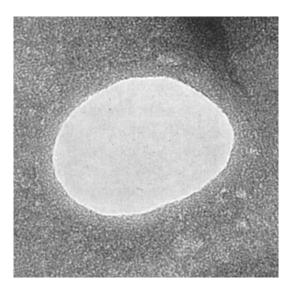
Bright line inside hole (objective lens too weak)

Near focus

Over focus

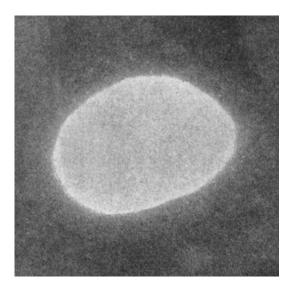
Bright line inside carbon film (objective lens too strong)

Hole in a Thin Carbon Film



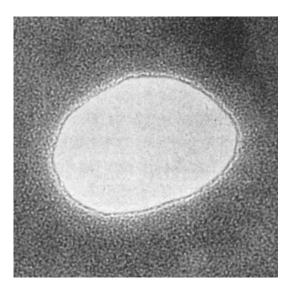
Under focus

Hole in a Thin Carbon Film



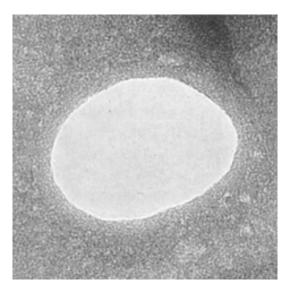
Near focus

Hole in a Thin Carbon Film



Over focus

Hole in a Thin Carbon Film

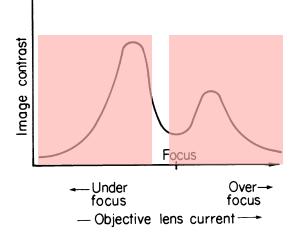


Focus series Animation



Rules of Thumb to Live (and Die) By:

- 1. The higher the magnification, the more accurately the image must be focused
- 2. High contrast does NOT mean you have achieved optimum focus



From Meek, 2nd ed., Fig. 5.3, p.100



- ★ Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
- \star Choice of magnification
- \Rightarrow Focusing
 - Magnification calibration
 - Resolution tests

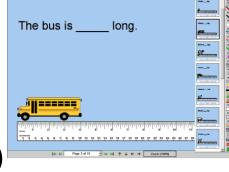


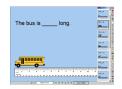
- Image intensifiers/TV displays Microscope maintenance
- Photography (analog and digital)



- ★ Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
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I.E.6 Magnification Calibration

Why be concerned?

~ 2-5% uncertainty in **nominal** magnification settings in modern TEMs

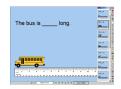
Must have an **independent** calibration of image magnification to measure specimen dimensions accurately

Why are nominal settings uncertain?

TEM lenses suffer from **hysteresis** and must be **normalized** to improve reproducibility (p. 32 of lecture notes), but there is no guarantee that actual image magnifications will precisely match the nominal settings.

What must be done?

Record images of **calibration standards** (specimens with known dimensions or spacings)





I.E.6 Magnification Calibration

Why be concerned?

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What must be done?

Record images of **calibration standards** (specimens with known dimensions or spacings)

Each specimen holder used must be calibrated separately





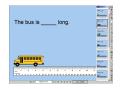
I.E.6 Magnification Calibration

Primary Calibration Standards

Polystyrene Latex Spheres

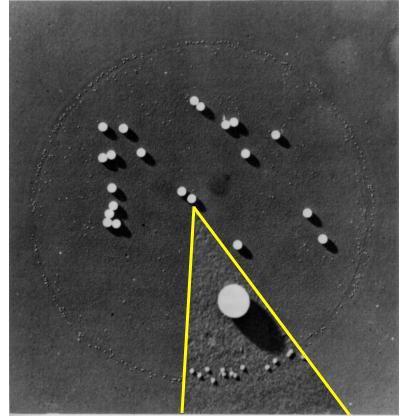
Diffraction Grating Replicas

Crystalline Specimens



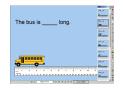
I.E.6 Magnification Calibration I.E.6.a Polystyrene Latex Spheres





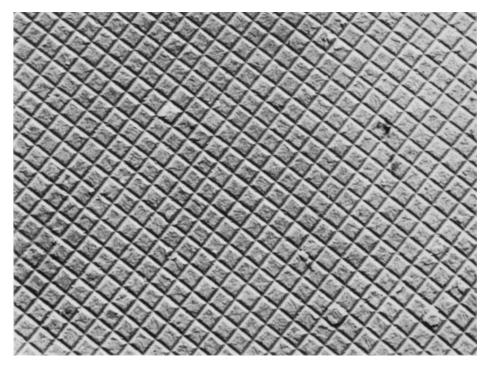
From Hall, Fig. 10.65, p.359

- Relatively uniform (come in various sizes ≥ 100 nm diameter)
- Serve as internal calibration standards (mix with specimen sample and image together)
- Accuracy only ~5-10%
- Tedious: need to measure large numbers of spheres



I.E.6 Magnification Calibration I.E.6.b Diffraction Grating Replicas

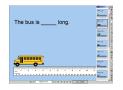




Replica of Cross-ruled Diffraction Grating (2160 lines/mm)

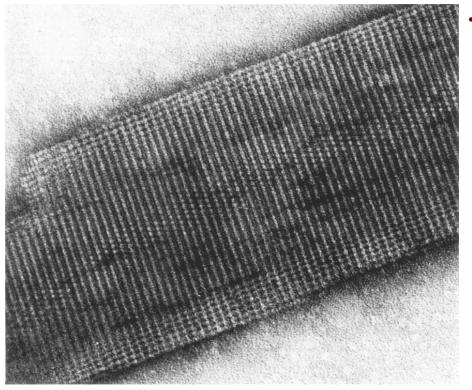
From Agar, Fig. 5.9, p. 162

Convenient calibration of low magnification TEM settings (~5,000-20,000X)



I.E.6 Magnification Calibration I.E.6.c Crystalline Specimens



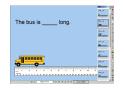


Thin crystal of beef liver catalase, negativelystained with ammonium molybdate

Periodicities: 8.75 x 3.43 nm

From Agar, Fig. 5.10, p. 163

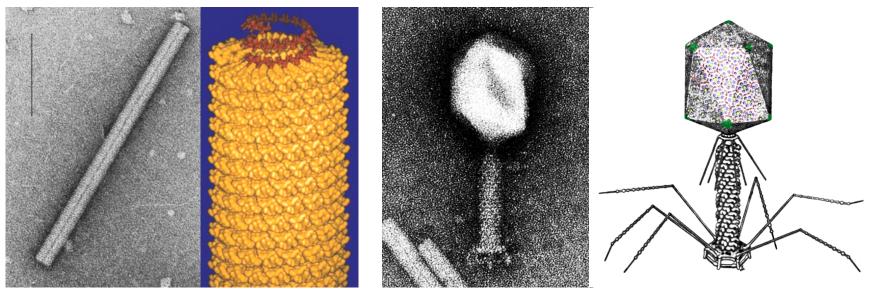
Convenient calibration of medium magnification TEM settings (~20,000-100,000X)



I.E.6 Magnification Calibration I.E.6.c Crystalline Specimens



Several good internal calibration standards for biological specimens



Tobacco mosaic virus (2.3 nm axial spacing)

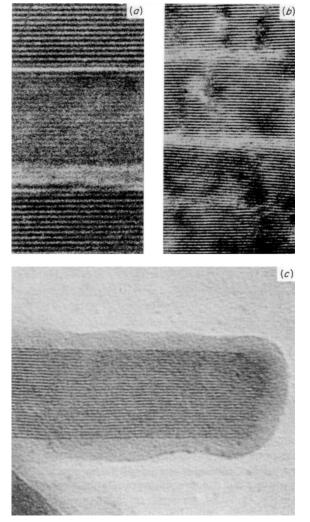
T4 bacteriophage tails (3.9 nm axial spacing)



I.E.6 Magnification Calibration I.E.6.c Crystalline Specimens



Calibration standards for high magnifications (>100,000X)



From Meek 1st ed., Fig. 12.6, p. 334

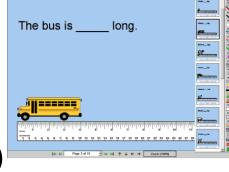
- (a) Cu-phthalocyanine (0.98 and 1.26 nm lattice spacings
- (b) K-chloroplatinate (**0.699 nm** spacing; but very susceptible to beam damage)

(c) Pt-phthalocyanine (1.25 nm spacing)



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I.E.7 Resolution Tests

Recall: Resolving power of the TEM

Defined as the best possible performance as limited by the small aperture of objective lens

How does one check microscope performance?

- Record micrographs of suitable test specimens
- Measure actual resolution achieved in the recorded image

I.E.7 Resolution Tests

Primary Resolution Tests

Point Separation (p.100 in lecture notes)

Lattice Resolution

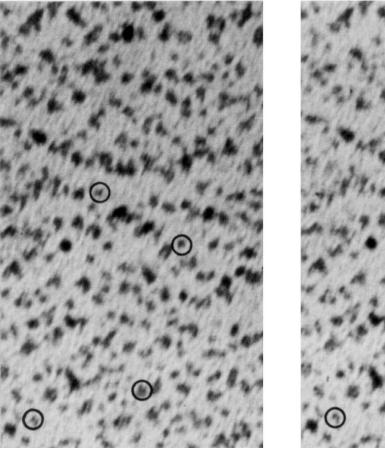
I.E.7 Resolution Tests

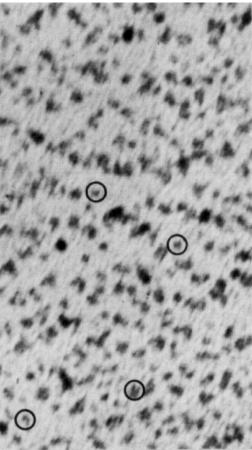
I.E.7.a Point Separation Test Point Separation Resolution Measurement

Platinum-iridium on carbon substrate

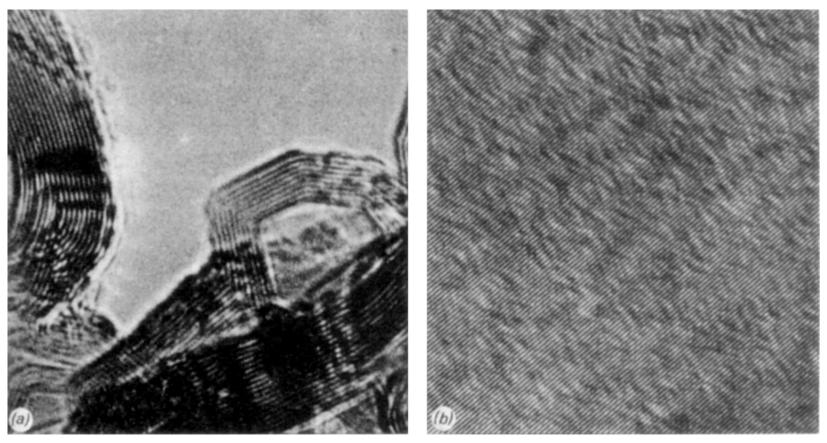
Separation of < 1.0 mm for pairs of particles, imaged at ~ 2 million times, indicates resolving power of better than 5 Å

Two micrographs needed to ensure that random phase granularity is not measured by mistake for actual particles





I.E.7 Resolution Tests I.E.7.b Lattice Resolution Test Crystal Lattice Spacings



Graphitized carbon (lattice spacing = 3.4 Å)

Gold foil (lattice spacing = 2.04 Å)

From Meek 2nd ed., Fig. 13.8, p. 317



- ★ Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
- ★ Choice of magnification
- ★ Focusing
- \star Magnification calibration
- ☆ Resolution tests
 - Image intensifiers/TV displays
 - Microscope maintenance
 - Photography (analog and digital)



- \star Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
- \star Choice of magnification
- ★ Focusing
- \star Magnification calibration
- ★ Resolution tests
- Image intensifiers/TV displays (pp.102-103, lecture notes) Microscope maintenance Photography (analog and digital)

I.E.8 Image Intensifier / TV Displays

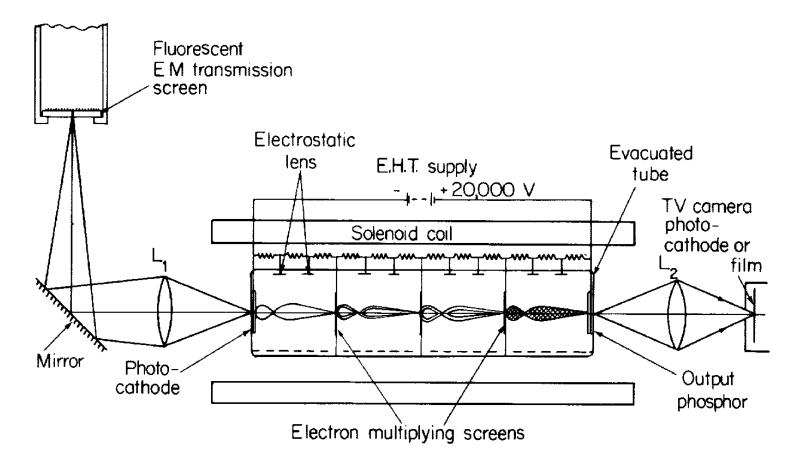


Diagram of TEM image intensifier developed by GEC-AEI Ltd..

From Meek 1st ed., Fig. 15.10, p. 401



- \star Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
- \star Choice of magnification
- ★ Focusing
- \star Magnification calibration
- ★ Resolution tests
- Image intensifiers/TV displays (pp.102-103, lecture notes) Microscope maintenance Photography (analog and digital)



- ★ Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
- \star Choice of magnification
- ★ Focusing
- \star Magnification calibration
- ★ Resolution tests
- ★ Image intensifiers/TV displays
- Microscope maintenance (p. 103 lecture notes) Photography (analog and digital)

BOTTOM LINE: Pay big bucks for a service contract

- ★ Choice of accelerating voltage
- ★ Choice of apertures
- ★ Specimen stage/holder
- \star Choice of magnification
- ★ Focusing
- \star Magnification calibration
- ★ Resolution tests
- ★ Image intensifiers/TV displays
- ★ Microscope maintenance
- \star Photography (analog and digital)



I.E.10 Photography (Film)

Photographic process **Optical density** Density related to exposure Density/exposure curves Contrast Speed (sensitivity) of electron emulsion Electron range in emulsion Number of grains per electron Graininess Resolution (image spread - electron diffusion)

I.E.10 Photography (Film)

☆ Photographic process **Optical density** Density related to exposure Density/exposure curves Contrast Speed (sensitivity) of electron emulsion Electron range in emulsion Number of grains per electron Graininess **Resolution** (image spread - electron diffusion)

I.E.10 Photography (Film) What's the Bottom Line?

Complete, faithful, and permanent record of details contained in the electron image

With film images in the TEM, the goal is to: Maximize Density Enhance Contrast Reduce Noise

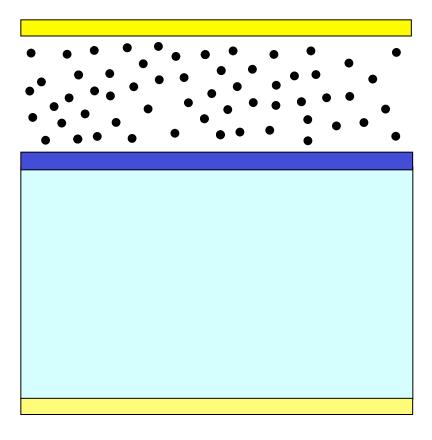
I.E.10 Photography (Film)

KEY CONCEPT

Photographic emulsions respond <u>differently</u> to <u>electrons</u> and <u>photons</u>

Electrons: single-hit process
Photons: multiple-hit process

I.E.10 Photography (Film) I.E.10.a The Photographic Process Structure of the Photographic Film



Hard gelatin-protective coat

Ag halide crystals

Substrate-adhesive

Emulsion support

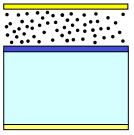
Anti-curl layer

I.E.10 Photography (Film)

I.E.10.a The Photographic Process

What happens when electrons or photons hit an emulsion?

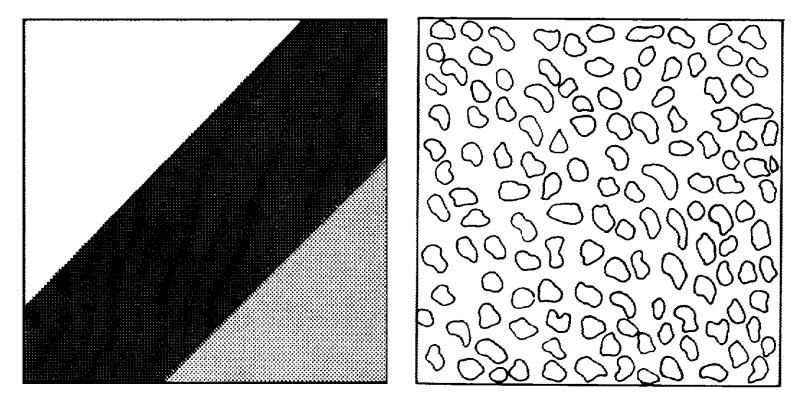
Ag atom **specks** form in the Ag-halide crystal



Photons: Required energy is ~ 30eV 10 photons needed (each photon has ~2.5eV)

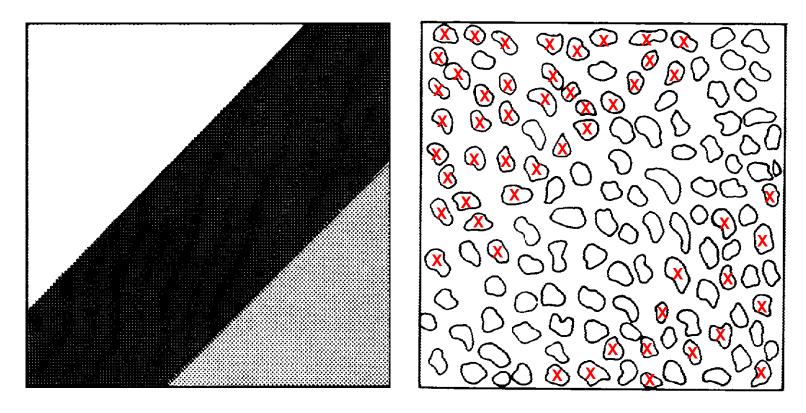
Electrons: Required energy is ~ 500eV 1 e⁻ in a 100keV beam more than sufficient !!!

Exposure of film to e⁻ is a **single-hit** process

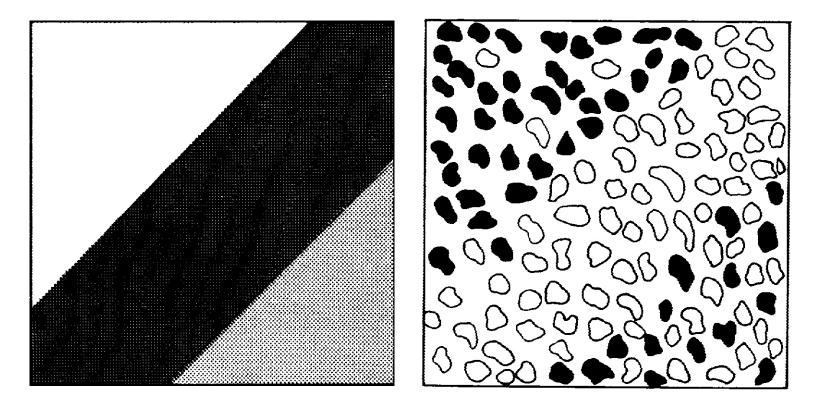


Electron image (Very high mag and small field of view)

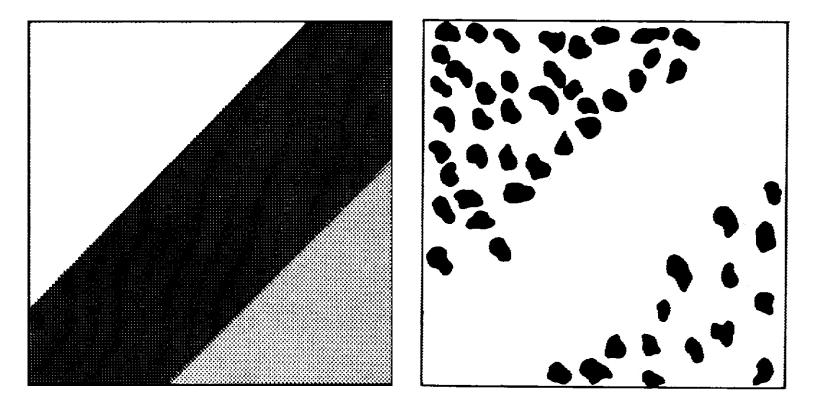
Unexposed emulsion (Area same as image on left)



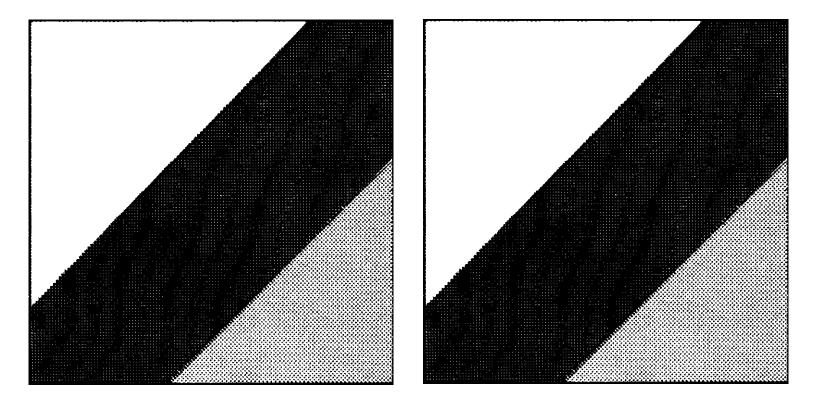
Electron image (Very high mag and small field of view) Exposed but unprocessed emulsion



Electron image (Very high mag and small field of view) Developed but unfixed emulsion



Electron image (Very high mag and small field of view) Final photographic image ("negative") after fixation



Electron image (Very high mag and small field of view) Electron image

I.E.10 Photography (Film)





I.E.10 Photography (Film)

I.E.10.b Optical Density (OD) of the Processed Emulsion

Optical Density:

Quantitative measure of how <u>black</u> the photographic emulsion gets when exposed to radiation

Definition:

$$OD = \log_{10}(I_o / T)$$

 I_o = intensity of **incident** radiation

T = intensity of **transmitted** radiation

I.E.10 Photography (Film)

I.E.10.b Optical Density (OD) of the Processed Emulsion

OD: quantitative measure of blackening of the emulsion

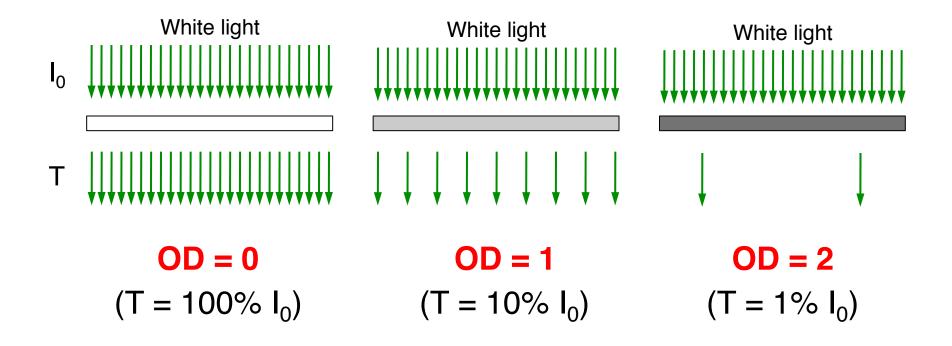
 $OD = \log_{10}(I_o / T)$

EXAMPLES:

OD = 1.0 for any portion of the photographic negative that transmits 10% of the incident light (T = 0.1)

OD = 2.0 for any portion that transmits only 1% of the incident light (T = 0.01)

I.E.10 Photography (Film) I.E.10.b Optical Density (OD) of the Processed Emulsion $OD = \log_{10}(I_o/T)$



I.E.10 Photography (Film)

- ★ Photographic process
- ★ Optical density
- Density related to exposure lecture notes p.106
- ☆ Density/exposure curves lecture notes pp.106-107
- ☆ Contrast lecture notes p.107
- ☆ Speed (sensitivity) of electron emulsion lecture notes p.108
- Electron range in emulsion lecture notes pp.108-109
 Number of grains per electron
 Graininess

Resolution (image spread - electron diffusion)

I.E.10 Photography (Film)

- ★ Photographic process
- ★ Optical density
- ★ Density related to exposure
- ★ Density/exposure curves
- ★ Contrast
- ★ Speed (sensitivity) of electron emulsion
- \star Electron range in emulsion
- Number of grains per electron
 Graininess
 Resolution (image spread electron diffusion)



I.E.10 Photography (Film) I.E.10.h Number of Grains / Electron



Each **electron** in the image likely hits **more** than 1 Ag halide crystal as it passes into and through the emulsion

~1 halide crystal for each 2 µm of emulsion thickness

~ 10 grains per electron track for 20 μ m thick emulsion

Each **electron** hits and passes through **several** halide crystals, losing some energy in each crystal

I.E.10 Photography (Film)

- ★ Photographic process
- ★ Optical density
- ★ Density related to exposure
- ★ Density/exposure curves
- ★ Contrast
- ★ Speed (sensitivity) of electron emulsion
- \star Electron range in emulsion
- Number of grains per electron
 Graininess
 Resolution (image spread electron diffusion)



I.E.10 Photography (Film)

- ★ Photographic process
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- \star Electron range in emulsion
- ★ Number of grains per electron
- ☆ Graininess lecture notes pp.109-110

Resolution (image spread - electron diffusion)



Statistical phenomenon caused by "electron noise" Random distribution of e⁻ "particles" in the beam

Graininess:

- NOT a defect in the photographic emulsion
- Product of two random processes



Graininess is the product of 2 random processes

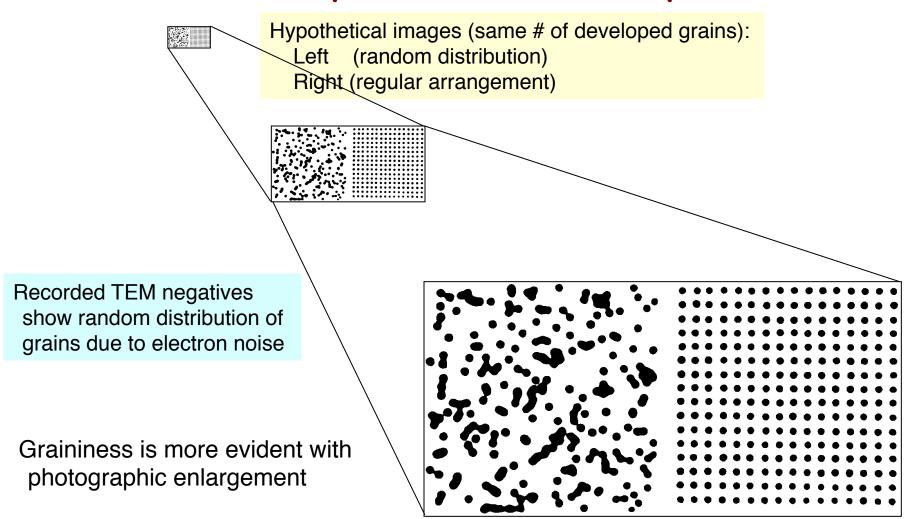
1. Random arrival of electrons at the film

Over **large** areas of an electron image, radiation appears **uniform** Over **small** regions, radiation distribution is **NOT uniform**

2. Granularity in the emulsion



Graininess is the product of 2 random processes



From Agar, Fig. 7.6, p. 201



Graininess is the product of 2 random processes

1. Random arrival of electrons at the film

Over **large** areas of an electron image, radiation appears **uniform** Over **small** regions, radiation distribution is **NOT uniform**

2. Granularity in the **emulsion**

Results from variations in:

- # of halide crystals hit by each electron in the image
- Area over which exposed crystal grows during development

Graininess in TEM images mainly caused by #1



Ways to Reduce Graininess

- To increase *S*/*N*, **need to increase "exposure"**
- 1. Use more electrons when recording images ("routine" TEM)
- 2. Use **image processing**: average many images together (effectively increases the # of electrons per image point)

3. Use film development strategies

Have minimal effect on e- noise but can reduce grainy appearance

I.E.10 Photography (Film)

- ★ Photographic process
- ★ Optical density
- ★ Density related to exposure
- ★ Density/exposure curves
- ★ Contrast
- ★ Speed (sensitivity) of electron emulsion
- \star Electron range in emulsion
- ★ Number of grains per electron
- ☆ Graininess lecture notes pp.109-110

Resolution (image spread - electron diffusion)

I.E.10 Photography (Film)

- ★ Photographic process
- ★ Optical density
- ★ Density related to exposure
- ★ Density/exposure curves
- ★ Contrast
- ★ Speed (sensitivity) of electron emulsion
- \star Electron range in emulsion
- ★ Number of grains per electron
- ★ Graininess
- \Rightarrow **Resolution** (image spread electron diffusion)

I.E.10 Photography (Film) I.E.10.j Resolution - Image Spread Emulsions Have Limited Resolution



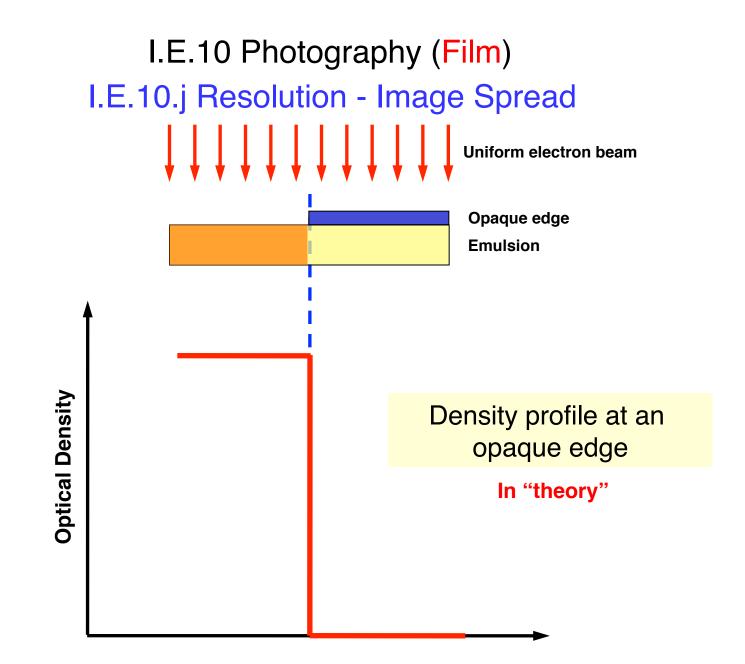
Emuisions flave limited Resolution

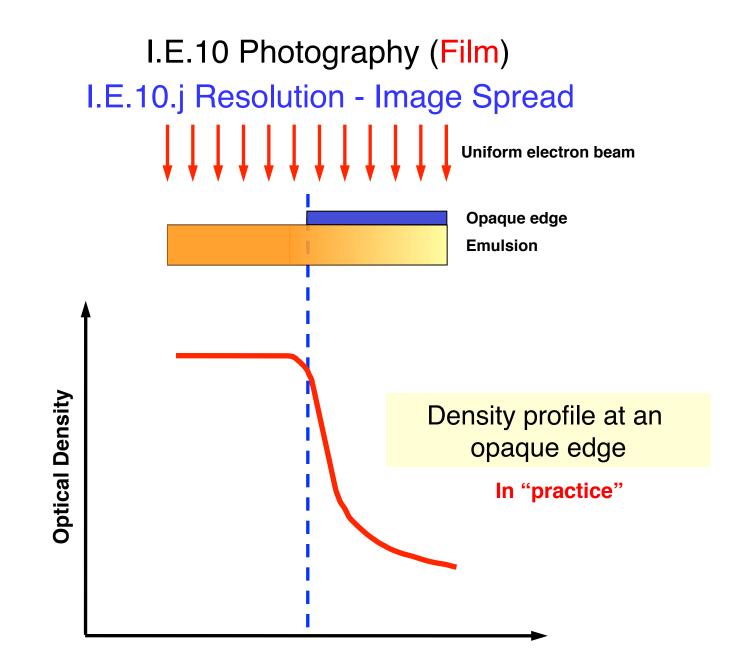
Electron tracks through the emulsion don't follow straight lines perpendicular to the surface

Electrons **scatter away** from the incident direction by interacting with atoms in the Ag halide crystals

Sideways scatter of electrons in the emulsion is called "electron diffusion"

.: Image details are spread out and contrast is reduced





I.E.10 Photography (Film) I.E.10.j Resolution - Image Spread

What's the Bottom Line?

Resolution in the FINAL (*i.e.* **recorded**) image is always **POORER THAN** the resolution achieved in the **electron** image

The solution?

Use magnification high enough to make sure details in the electron image are captured by the emulsion

But not too high!!! (restricted field of view, excessive damage, etc.)



I.E.10 Photography (Film) I.E.10.j Resolution - Image Spread

Seems like everything affects resolution

Size of lens aperture (diffraction effects)

Lens aberrations (spherical, chromatic, asymmetry)

Defocus effects (phase contrast Fresnel fringes)

Emulsion grain size

Electron diffusion in emulsion

And yes, you can expect some more...



§ I: The Microscope

I.E Operation of the TEM

I.E.10 Photography (Film)
 I.E.11 Digital Photography (CCD)

I.E OPERATION OF THE TEM

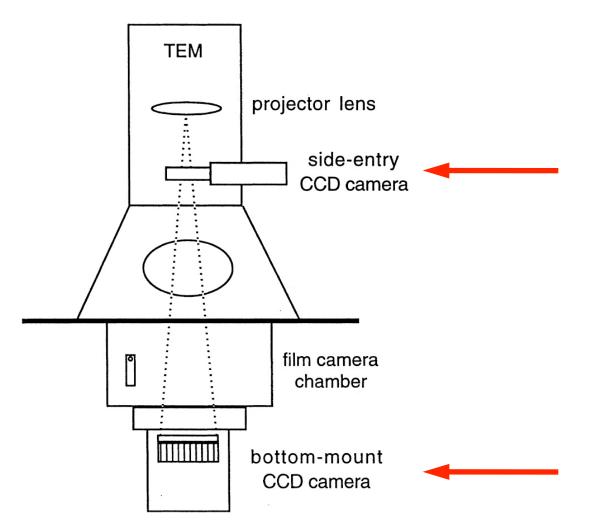
I.E.11 Digital Photography (CCD)

CCD Detectors/Cameras

CCD = Charge Coupled Device

I.E OPERATION OF THE TEM

I.E.11 Digital Photography (CCD)



I.E.11 Digital Photography (CCD) Quotable Quotes



Digital

Film

I.E.11 Digital Photography (CCD) Quotable Quotes

...likely consequence of these trends is that TEM film cameras and EM photographic darkrooms will ultimately be relegated to technical museums. As a practical example, this entire paper was prepared without any use of silver-halide-based photographic emulsions, and its authors have no intention of ever stepping into a photographic darkroom again.

Krivanek & Mooney Ultramicrosc. 49[1993]95

Direct digital recording with slow-scan CCD cameras [...] provides a significant advantage over film such as higher sensitivity and dynamic range. [...] performance characteristics [...] challenge the dominance of film as the recording medium for many scientific imaging requirements. [...] These cameras are revolutionizing the way microscopists think about addressing problems of data acquisition ...

Fan & Ellisman Ultramicrosc. 52[1993]21

...data acquisition in the electron microscope will increasingly be with electronic detectors. ...CCDs are <u>not yet suitable</u> for high resolution imaging.

Farugi, Henderson, & Subramaniam Ultramicrosc. 75[1999]235

...digital imaging systems appear to be <u>poised to replace film</u> in TEM. ...a <u>much larger array size will be needed</u> to match the large field of view and the great details typically contained in a piece of TEM film. Fan et al., Ultramicrosc. 84[2000]75

CCDs are *increasingly replacing photographic film* as the recording medium of choice in electron microscopy. Meyer et al., Ultramicrosc. 85[2000]9

... traditional and still the most common recording medium is photographic film ...

Koeck Microsc. Res. & Tech. 49[2000]217

So where do we stand today?

Film has already been replaced by CCD cameras in many applications

Film and CCDs are in the process of losing the next "battle" against DDDs

$DDD = \underline{D}irect \underline{D}etection \underline{D}evice$

(measures electron events directly)

Advantages of Recording Images by CCD

- Immediate image access (no tedious film development)
- Much larger dynamic range than film
- Strict linear response with electron dose
- Amenable to all types of **automated** experiments

Auto-focusing and astigmatism correction
Auto-imaging
Electron tomography
Electron holography
Protein electron crystallography
Telemicroscopy
etc. etc.

Disadvantages of Recording Images by CCD

- **Poorer "pixel" resolution** (15 μ m vs. ~ 5-10 μ m for film)
- Limited number of pixels, hence small field of view

Affordable CCDs have 1k² or 2k² pixel arrays

Film is comparable to 16,000 by 20,000 pixels

- High upfront cost (~ \$200K for high quality 4k² camera)

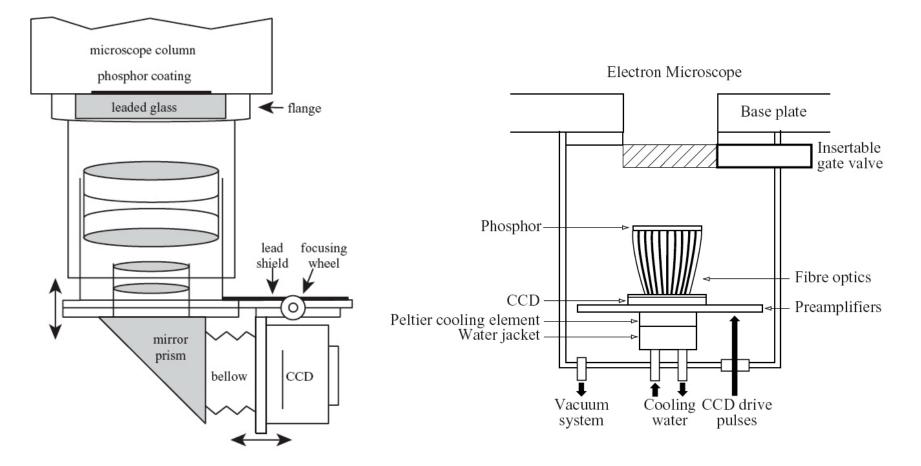
CCD Topics (Woulda, Coulda, Shoulda...)

- CCD design (lens or fiber optic coupling)
- Mount in TEM (in-line or retractable or off-line)
- Scintillators (YAG; P43 GOS; red P20 phosphor)
- Dark current; gain normalization; readout noise; Peltier cooling
- Pixel size; pixel resolution (PSF: point spread function); pixel binning
- MTF (modulation transfer function of camera)
- Nyquist frequency/sampling
- DQE (detective quantum efficiency of electron detection)
- Dynamic range
- Distortion

I.E.11 Digital Photography (CCD) Two Basic CCD Designs

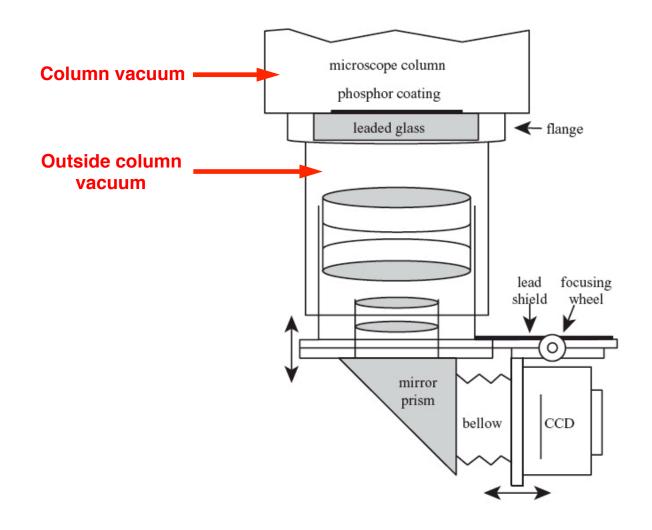
Lens-Coupled

Fiber Optic-Coupled



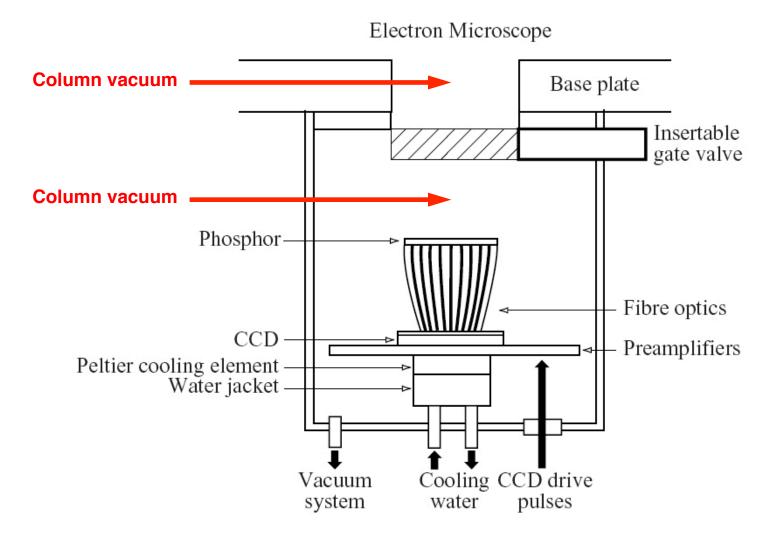
From Faruqi & Andrews [1997], p.234

I.E.11 Digital Photography (CCD) Lens-Coupled CCD



From Fan & Ellisman [1993], p.22

I.E.11 Digital Photography (CCD) Fiber Optic-Coupled CCD

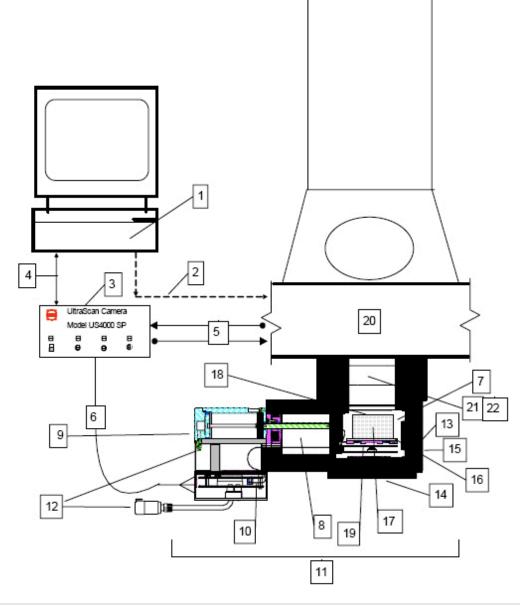


From Faruqi & Andrews [1997], p.234



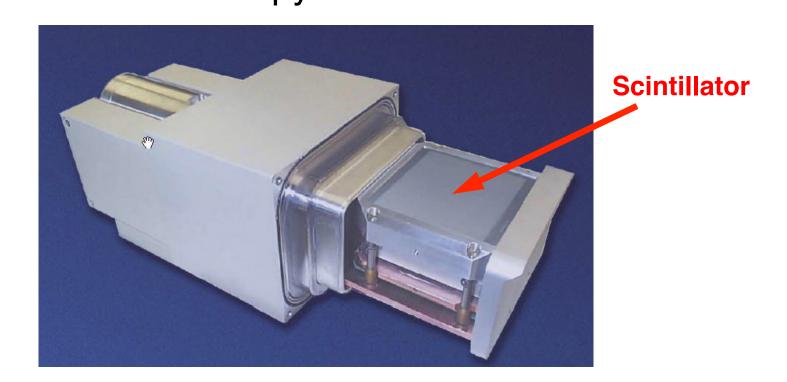
UltraScan System

- 1. Computer
- 2. TEM-interface cable (optional)
- 3. Controller
- 4. Camera-interface cable
- 5. Shutter-interface cable
- 6. Camera-interface cable
- 7. UltraScan vacuum chamber
- 8. Drive assembly
- 9. Pneumatic inlets
- 10. Preamp assembly
- 11. UltraScan camera module
- 12. Coolant connections
- 13. X-ray shielding
- 14. Blanking flange
- 15. Accessory port
- 16. Charge-coupled device
- 17. Fiber optic plate
- 18. Electron scintillator
- 19. Peltier cooler
- 20. Electron microscope
- 21. Microscope vacuum
- 22. UltraScan flange



From Gatan, Inc. Ultrascan 4000 Userguide, p.48

I.E.11 Digital Photography (CCD) Microscopy with a CCD



CCDs like the 16 megapixel Gatan Ultrascan[™] (4080 x 4080 15 μm pixels) can produce high quality digital images in the TEM

http://www.gatan.com/imaging/ultrascan.html